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## Partial Purification of Extracellular Lipase from Soil Fungal Isolate

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### ABSTRACT

Lipases are biotechnologically vital cluster of enzymes reported to be produced by many species of animals, plants, bacteria, yeasts and fungi. Lipases of microbial origin that are employed in the detergent, dairy, cosmetic, food, and tanning industries are mostly engaging. The present study was aimed at partial purification of lipase produced from soil fungal isolate. Hyper producer strain was selected by growing fungal isolate onto the tributyrin agar media plates. Extracellular lipase was produced by inoculating the isolated strain in fermentation broth followed by incubation at 28°C for 5 days at 120rpm. Mycelium free culture supernatant obtained after centrifugation of fungus broth was used for purification study. Among the ammonium sulphate precipitates, 60% fractionate showed the highest lipase activity and protein content, and on further dialysis the lipase activity was increased. 2.78 fold purification was obtained by 60% ammonium sulphate dialyzed precipitation which is higher than the fold purification of crude lysate.

**Keywords:** Crude lysate, fungi, lipases, partial purification.

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## INTRODUCTION

Lipases (triacylglycerol acylhydrolase, EC 3.1.1.3) are a biotechnologically vital cluster of enzymes that act on the carboxyl organic compound bonds present in triacylglycerols and liberate fatty acids and alcohol.<sup>1</sup>

Lipases are reported to be produced by many species of animals, plants, bacteria, yeasts and fungi.<sup>2</sup> Among these, microbial lipases have gained wide industrial applications in view of high yields, rapid growth of microorganisms and ease of genetic manipulation.<sup>3</sup>

Lipases chemical action potentials are monstrous. Lipases of microbial origin that are employed in the detergent, dairy, cosmetic, food, and tanning industries are mostly engaging. In view of their recent and potential applications, lipases are thought to be a promising category of commercial enzymes.<sup>4</sup>

The purification of lipase from culture filtrate was reported by SDS-PAGE and disc-PAGE.<sup>5</sup> The purification included gel filtration, SP-Sephadex and isoelectric focusing and obtain improved yield was 38%. 31 KDa molecular weight of protein was determined by SDS-PAGE. 134 fold purification of extracellular lipase present in production broth was estimated through ammonium sulphate precipitation, gel permeation and ion exchange chromatography with yield of 46%.<sup>6</sup>

33.7% yield was obtained by partial ammonium sulphate precipitation and ion exchange chromatography. 90 KDa molecular weight of lipase was determined and optimum activity at pH 7 and 40°C. By keeping immense applications of lipase in our mind we have produced lipase by soil fungi followed by ammonium sulphate precipitation, dialysis and acetone precipitation.<sup>7</sup>

## MATERIALS AND METHOD

All chemicals and glassware's were used from Rankem, Sigma Aldirich, SRL, TM media, Hi media, Borosil and Riviera.

### Sample collection

The soil sample was collected from petrol pump, Newai (Raj). Soil sample was collected with sterile spatula in a sterile plastic bag and stored in refrigerator at 4°C until use.

### Isolation of lipolytic fungi

Isolation of lipolytic fungi was done by serial dilution and spread plate method. PDA plates were incubated at 28°C for 5-7 days in inverting condition.<sup>8</sup>

### Screening of lipase producing fungi

The pure fungal isolates were screened onto tributyrin agar (TBA) plates. TBA plates were point inoculated with fungal isolates and were incubated at 28°C for 4-5 days in inverted

condition. After incubation plates were analyzed for appearance of clear zone around the colonies.<sup>9</sup>

### **Production of extracellular lipase from submerged fermentation**

Highest zone of clearance produced strain was selected and grown in culture media. One ml of spore suspension was inoculated in 100 ml of production medium which was prepared by dissolving (4 g of peptone, 0.1g of MgSO<sub>4</sub>.7H<sub>2</sub>O, 0.1 g of KH<sub>2</sub>PO<sub>4</sub>, 0.1 g of (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, 0.5 g of Sucrose, 1 ml of olive oil) under aseptic conditions and pH was adjusted to 6.5±0.2. Inoculated flasks were kept in shaking incubator at 120 rpm, 28°C for 6-7 days.<sup>10</sup>

### **Preparation of crude protein lysate**

Fermentation broth from inoculated flask was filtered by using Whatmann filter paper No. 1 to remove mycelium network. Filtrate was centrifuged at 10,000 rpm for 10 min. at 4°C. Pellet was discarded and supernatant was considered as crude protein lysate for determination of and lipase activity and protein content.<sup>11</sup>

### **Partial purification of extracellular lipase**

#### **Acetone precipitation**

The crude enzyme lysate was subjected to 60% (v/v) fractionate by adding chilled acetone and mixed with continuous stirring and left overnight at 4°C. The ensuing acetone precipitate was centrifuged at 10,000 rpm for 20 min at 4°C. This resulted in pellet formation and was dissolved in minimal amount of tris HCl buffer (0.02 M, pH7). The lipase activity and protein content was determined in pellet and supernatant both.

#### **Ammonium sulfate precipitation**

The crude enzyme lysate was subjected to different fractionate (60%, 80% and 90%) of salt saturation by adding solid ammonium sulfate calculated through given formula. Resultant precipitates of each fractionate was centrifuged at 10,000 rpm for 20 min at 4°C and dissolved in minimum amount of tris HCl buffer (0.02 M, pH7) further dialyzed extensively against the same buffer overnight at 4°C. The lipase activity was calculated after each fractionate in both pellet and supernatant.

$$\text{Grams of (NH}_4\text{)}_2\text{SO}_4 = \frac{533 \times (S_2 - S_1)}{100 - 0.3 \times S_2}$$

Where, S<sub>1</sub> is initial percent salt concentration and S<sub>2</sub> is percent salt saturation we want to obtain.

This equation is based on assumption that 100% saturation is equal to 4.05 M (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>.

In order to determine, how much pure protein we get as compared to crude lysate fold purification and % yield were calculated using given formulas.

$$\text{Fold purification} = \frac{\text{Specific Activity of each fraction}}{\text{Specific Activity of crude protein lysate}}$$

$$\% \text{ Yield} = \frac{\text{Total enzyme activity of each fraction}}{\text{Total enzyme activity of crude protein lysate}} \times 100$$

### Dialysis

Fraction of ammonium sulphate precipitation was subjected to dialysis to remove the residual impurities from the purified enzyme mixture. Dialysis membrane (cut off value 10KDa) was used. One end of the membrane was tied up with thread and enzyme was poured into the dialysis membrane. The other end of membrane was tied securely. The membrane was put into the 200 ml of tris HCL buffer (0.02 M, pH 7) and was kept on magnetic stirrer for 2 hours at room temperature. The buffer was replaced after 2 hours and it was left overnight. The enzyme activity and protein content was checked.

### RESULTS AND DISCUSSION

The partial purification process and results are summarized in Table 1. Crude lysate was fractionate and dialyzed to get enzyme activity. Lipase activity and protein content was checked in each fractionate. 60% ammonium sulfate precipitation shown highest lipase activity among all different fractionates. 60% ammonium sulfate was followed by dialysis which resulted in increase of lipase activity (15.48±0.11 IU/ml/min). 2.78 fold purification was obtained by the purification process of 60% dialysate.

**Table 1: Purification table showing ammonium sulphate precipitation, dialysis and acetone precipitation of crude enzyme extract.**

Steps	Vol (ml)	Lipase activity IU/ml/min	Protein contents mg/ml	Total activity Units * ml	Total protein content * ml	Specific activity U/mg	Fold purification	% yield
Crude protein lysate	100	6.89±0.09	16.37	689	1637	0.41±0.005	1	100
<b>Ammonium sulphate precipitation</b>								
60% fraction	114	7.03±0.73	22.08	801.42	2517.12	0.32±0.05	0.78	116.31
60% supernatant	112	1.93±0.26	8.22	216.16	920.64	0.23±0.03	0.56	31.51
80% fraction	117	3.62±0.52	21.58	423.54	2524.86	0.16±0.02	0.39	61.47
80% supernatant	115	2.62±0.82	4.98	301.3	572.7	0.52±0.17	1.26	43.73
90% fraction	119	6.29±0.16	7.44	784.51	885.36	0.84±0.03	2.04	113.86
90% supernatant	117	4.09±0.61	0.42	478.53	49.37	1.16±0.12	2.82	69.45
<b>Dialysis</b>								
Dialysis (60% fraction)	8.6	15.48±0.11	13.53	133.12	116.35	1.14±0.02	2.78	19.32
Dialysis (80% fraction)	4.8	10.25±0.39	12.59	49.2	60.43	0.81±0.01	1.97	7.14
Dialysis (90% fraction)	0.8	5.61±0.24	4.11	4.48	3.28	1.36±0.10	3.31	0.65
<b>Acetone precipitation</b>								

60% fraction	100	6.72±0.26	9.97	672	997	0.67±0.02	1.63	144.70
60% supernatant	98	0.61±0.40	8.43	59.78	826.14	0.04±0.02	0.09	119.90

Dialysis of 60%, 80% and 90% leads to an increase in specific activity and fold purification. 60% and 90% fractionate shown more % yield then crude protein lysate. The 90% ammonium sulphate fractionate show highest specific activity (0.84±0.03 U/mg) and 2.04 fold purification than crude enzyme lysate, and further dialysis increased activity (1.36±0.10 U/mg) and 3.31 fold purification. 1.63 fold purification was obtained by 60% acetone fraction precipitation higher than crude protein lysate.

In our experiment, we used ammonium sulfate for the fractionation step purification because of its solubility in water is higher due to its ionic character; therefore proteins are salted out by precipitation and called “salting out”. Also used because it has ability to become more hydrated compared to non polar molecules and precipitate out of the solution. Exact amount of ammonium sulfate addition in the solution leads to precipitation of selectively protein while some remains soluble in solution.

Highest lipase activity was observed in all fractionate then in crude enzyme lysate. The highest fraction was F60 having value 12,283.33 U/mg protein and was used for further characterization process.<sup>12</sup> Maximum specific activity (39.70 U/mg protein) was reported by ammonium sulfate precipitation then crude lysate.<sup>13</sup> Similar to our result, more lipase activity was obtained after 60-80% dialyzed and obtained 2.9 fold purification.<sup>14</sup>

After the enzyme produced in production media it was subjected to ammonium sulphate precipitation for purification of proteins. It was obvious from the result that 60% supernatant ammonium sulphate exhibited more lipase activity then the crude lysate.<sup>15</sup> 25.4 U mL<sup>-1</sup> extracellular lipase was obtained after 80% ammonium sulphate precipitation.<sup>16</sup>

## CONCLUSION

The main aimed of study was purification of extracellular lipase isolated from fungi of petrol pump soil sample. The 60% ammonium sulphate fractionates was acted as best inducer because it shows highest lipase activity (7.03±0.73 IU/ml/min) and 0.78 fold purification and increased on dialyzed 15.48±0.11 IU/ml/min with 2.78 fold purification. Column chromatography will be used in further research to obtain more purified lipase.

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